Increasing of IFNγ Expression in Transformed Cells of Balb/c Mice Due to Ethanol Extract Cocoa Beans (Theobroma cacao)

Peningkatan Jumlah Ekspresi IFNγ pada Mencit Balb/c yang Mengalami Transformasi Sel Akibat Pemberian Ekstrak Etanol Biji Kakao (Theobroma cacao)

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Abstract

As current technology cannot cure cancer completely, prevention becomes the main choice. To prevent the development of cell transformation into cancer cells, polyphenols that are widely found in cocoa beans (Theobroma cacao), is usable. IFNγ plays an important role in immunity against cancer. This study aims to see the ability of cocoa beans ethanol extract to increase the number of IFNγ expression in Balb/c mice undergoing cell transformation. This study used three groups Balb/c (n=4), namely K1 (normal control), K2 (negative control: injected with benzopyrene without ethanol extract of cocoa beans), and K3 (treatment: injected with benzopyrene, given 4mg/30gBW/po/day ethanol extract of cocoa beans). The mice were biopsied, and IFNγ expression was examined by immune histochemical method. The results showed that IFNγ expression increased significantly in K3. It can be concluded that ethanol extract of cocoa beans could increase IFNγ expression in Balb/c mice undergoing cell transformation.

Keyword: IFNγ, cells transformation, Theobroma cacao

INTRODUCTION

One cause of death in the world is cancer. In Indonesia, cancer is the seventh leading cause of death (5.7%).1 Of all cancers that occur in humans, head and neck cancer ranks 6th (3%). Most (48%) of head and neck cancers occur in the oral cavity, and 90% are squamous cell carcinoma of the oral cavity (SCC).2

Current technology is not able to cure cancer completely, while new cases continue to emerge. This causes the number of cancer patients including SCC continues to increase. Therefore, preventive measures are the first choice to prevent the rate of increase in the number of SCC patients.

Cancer prevention can be done at the stage of initiation, promotion, and cancer progression.3 The promotion phase transforms the transformed cells into cancer cells. This stage lasts for a long time, and some can experience regression. This provides an
opportunity to inhibit or prevent the occurrence of SCC. Polyphenols can be used to prevent the development of the transformation and preneoplastic cells into cancer cells.³ Polyphenols are found in many cocoa beans (Theobroma cacao) which are widely produced in plantations spread almost throughout Indonesia. Indonesian people used to call cocoa plants as cocoa.

IFNγ plays an important role in immunity against cancer.³ There has never been any research on the potential of ethanol extract of cocoa beans in increasing IFNγ expression in Balb/c mice undergoing cell transformation.

Based on the description above, research is needed to determine the ability of ethanol extract of cocoa beans to increase the amount of IFNγ expression in Balb/c mice undergoing cell transformation. This study aimed to see the ability of ethanol extract of cocoa beans to increase the amount of IFNγ expression in Balb/c mice undergoing cell transformation.

MATERIALS AND METHOD

This research was conducted at the Biochemical Laboratory and Electron Microscope Unit of the Faculty of Medicine, Airlangga University after being approved by the Ethics Commission of the Faculty of Dentistry, Airlangga University. This research was conducted with pure experiment with a completely random design. This research was an in vivo study using Balb/c mice models that have transformed. There were three groups in this study, namely K1 (normal control: without injection of benzopyrene, without ethanol extract of cocoa beans), K2 (negative control: injected with benzopyrene, without ethanol extract of cocoa beans), and K3 (treatment: injected with benzopyrene, given 4mg/30gBB /PO/day ethanol extract of cocoa beans). Four repetitions were carried out in each group so that the total sample was 12 mice. Mice had the following criteria such as male, healthy, age ± 2 months, and weight 20-30 grams. Previously, mice were adapted for one week, fed and drank ad libitum.

The animal model was made after being adapted for a week by exposing mice with 0.08 mg benzopyrene/0.04 ml of oleum olivarum in the oral cavity of the right buccal mucosa. The exposure was done by injection three times a week for three consecutive weeks. After that, it was left for two weeks.

The ethanol extract of cocoa beans was begun with the selection of cocoa pods, which were ripe cocoa pods from Forester/Lindak types obtained from Banjarsari Gardens, PTPN XII. After being picked and peeled, the cocoa beans were removed and cleaned. Then, it was dried by drying in the sun. The cocoa beans that have been dried were peeled and pollinated with a mesh particle size no. 25-35. A total of 3 kg of cocoa beans (Theobroma cacao), which have been pollinated were extracted by maceration with n-hexane solvent. At first, the cocoa bean powder was put into a maceration container, then n-hexane was added until the entire sample was submerged. The container was tightly closed and left for one day while stirring several times. The mixture was filtered and remastered by adding n-hexane to the pulp. The extraction process was carried out until the color of the solvent was clear. From this process, the low-fat cocoa pulp was produced. Furthermore, the extraction of polyphenols with 80% ethanol solvent in a ratio of 1:3 was done. The extraction was done by maceration method for three hours at room temperature while stirring every 15-20 minutes, then it was filtered. The results of the filter were evaporated using a vacuum evaporator to produce ethanol extracts of cocoa beans.⁶

The extracts were tested for the total phenolic content measured by the Folin-Ciocalteu method.⁷ The extract of 253.4 mg was dissolved with acetone: water (7:3). Then, the volume of up to 50 ml was adequate into a flask. 1 ml of sample solution was pipetted and put into a 50 ml flask and 20 ml of distilled water was added. A total of 2 ml of Folin-Ciocalteu reagent was added to the flask and left for 5 minutes. After that, 20 ml of 7% sodium carbonate were added to the mixture, and the volume was made to 50 ml with distilled water. The mixture was left for 90 minutes. The absorbance of the mixture was measured at 750 nm using a spectrophotometer. Standard curves were made using various concentrations of tannic acid. The results were expressed as percent tannic acid.⁸

The flavonoid content of the cocoa bean extract was determined by the aluminum chloride colorimetric assay method.⁹ An extract of 251.1 mg was dissolved with methanol as a solvent. The volume was up to 50 ml in a flask. An amount of 0.5 ml of the sample solution was pipetted and put in a 50 ml flask and then 20 ml of distilled water was added. 1.5 ml of 5% NaNO2 was left for 5 minutes. After 5 minutes, 10 ml of AlCl3 10% was added to make a volume of 50 ml with distilled water. The absorbance of the solution was measured at 510 nm. The standard curves are made using various concentrations of catechins. The results were expressed as percent catechins.
At the beginning of the seventh week, the experimental animals in the k3 group were given extracts at a dose of 4mg/30gBB/po/hrby using gastric sonde. The extract was administered for up to 4 weeks.

At the beginning of the 11th week, the right cheek mucosal tissue was taken by biopsy. Previously, the mice were anesthetized with ether, and then the mice were sacrificed. Biopsy tissue has been put into for formalin and used as an IFNγlevel examination by the immunohistochemical method.

The data obtained were presented descriptively, and the normality, homogeneity, and difference tests were carried out. Data analysis was performed with a computer program, the SPSS version 21.

RESULT

At the beginning of the 11th week, mice were biopsied and sacrificed. The results of measuring IFNγ expression can be seen in Table 1.

Table 1. Mean and standard deviation (SD) of the IFNγ expression

<table>
<thead>
<tr>
<th>Group</th>
<th>n</th>
<th>IFNγ (/625 μg) (mean± SD)</th>
</tr>
</thead>
<tbody>
<tr>
<td>K0</td>
<td>4</td>
<td>1.200 ± 0.135</td>
</tr>
<tr>
<td>K1</td>
<td>4</td>
<td>1.025 ± 0.125</td>
</tr>
<tr>
<td>K2</td>
<td>4</td>
<td>3.750 ± 1.452</td>
</tr>
</tbody>
</table>

Based on the results of the normality test with the Shapiro-Wilk test, the amount of IFNγ expressions was normally distributed. Homogeneity test with the Levene test shows that the data was not homogenous; therefore, a nonparametric statistical test was performed with the Kruskal-Wallis test to determine differences in all groups. The results showed that there was a significant difference in at least one group with p = 0.004 (<0.05), followed by the Mann-Whitney test to see which groups were different. The results can be seen in Table 2.

Based on Table 2, there was a significant difference between K0 group with K2 group, and K1 group with K2 group.

Table 2. The Mann-Whitney test results on the amount of IFNγ expressions

<table>
<thead>
<tr>
<th></th>
<th>K0</th>
<th>K1</th>
<th>K2</th>
</tr>
</thead>
<tbody>
<tr>
<td>K0</td>
<td></td>
<td>0.557</td>
<td>0.020*</td>
</tr>
<tr>
<td>K1</td>
<td>0.0021*</td>
<td></td>
<td></td>
</tr>
<tr>
<td>K2</td>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

*:significant

DISCUSSION

The increase in the prevalence of cancer including Oral Squamous Cell Caarcinoma (OSCC), is inseparable from the poor lifestyle of the community, among others due to smoking. Several things can cause cancer risk, but smoking is said to be the riskiest material to cause OSCC. This happens because cigarettes contain carcinogenic benzopyrene. Benzopyrene is also present in motorcycle exhaust fumes and smoked or baked foods. Therefore, in this study, the creation of a model of mice undergoing cell transformation was done by the induction of benzopyrene.

In addition, the modelling of mice undergoing cell transformation in this study was carried out by administering benzopyrene injection, which was a polycyclic aromatic hydrocarbon compound that had mutagenic and highly carcinogenic effects so that it was widely investigated. This compound can enter the body through the mouth, nose, and skin.

The results showed that the groups that were given ethanol extract of cocoa beans (K2) expressed a significantly higher amount of IFNγ compared to the normal control group (K0) and negative control (K1). This means that the ethanol extract of cocoa beans can increase IFNγ expression.

This happens because the ethanol extract of cocoa beans with their flavonoid content (catechin, proanthocyanidin, and anthocyanidin) is thought to increase IFNγexpression through the increase in macrophage cell activity through the activation of IKB Kinase (IKK) resulting in inactivation of IKβ. This causes NFκβ which was originally bound to IKβ to be active and translocated into the cell nucleus. This increases the activation of NFκβ responsive genes, which then increases cytokine production. In this case, it causes an increase in IL-1 and IFNγ cytokines. IL-1 activates Th and proliferates into Th1 /CD4 cells that produce IFNγ.

In this study, the normal control group also showed a small amount of IFNγexpression. This is possible because physiologically macrophages are also active in mice. In addition, at the time of the study, it was difficult to find sterile oral cavity of the mice. Based on the study results, it can be concluded that the ethanol extract of cocoa beans can increase IFNγ expression in Balb/c mice undergoing cell transformation.
REFERENCES

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