# THE EFFECT OF ETHANOLIC EXTRACT OF Syzygium cumini LEAVES ON THE GROWTH OF Streptococcus mutans

## PENGARUH EKSTRAK ETANOLIK DAUN JAMBLANG (Syzygium cumini) TERHADAP PERTUMBUHAN Streptococcus mutans

Suzanna Sungkar<sup>\*</sup>, Tetiana Haniastuti<sup>\*\*</sup>, Al. Supartinah<sup>\*\*\*</sup>, Dewi Agustina<sup>\*\*\*\*</sup>

\*Department of Pediatryc Dentistry Faculty of Dentistry Syiah Kuala University
\*Department of Oral Biology Faculty of Dentistry.Universitas Gadjah Mada
\*\*\* Department of Pediatryc Dentistry Faculty of Dentistry.Universitas Gadjah Mada
\*\*\* Department of Oral Medicine Faculty of Dentistry.Universitas Gadjah Mada
\*\*\* Tengku Tanoh Abee Street, Darussalam, Banda Aceh
Corresponding Email: suzannasungkar@unsyiah.ac.id

#### Abstract

Streptococcus mutans plays an important role in the pathogenesis of caries. This bacteria has virulence properties involve in the formation of biofilm on tooth surface. Due to its antibacterial effect, Jamblang leaf/Syzygium cumini L may be used as an agent to prevent caries. This study aimed to elucidate the effects of ethanolic extract of Syzygium cumini L on the growth of *S. mutans*. Analysis of active compounds was carried out using *thin layer chromatography* (TLC) and *Liquid Chromatograph-Mass Spectrography* (LC-MS). Samples of *S. mutans* were isolated from children's carious deciduous molar teeth. Growth test was done by dilusion technique. Aquadest was used as a negative control. TLC and LC-MS showed the presence of the flavonoid, tannin, and terpenoid *Kruskall-Wallis* test showed significant differences (p<0.05) among the groups, indicating that Jamblang leaves ethanolic extract decreased the growth of *S. mutans* at 22.5% of extract's concentration. *Syzygium cumini* L ethanolic extract reduces the growth of *S. mutans*. Concentration of 22.5% *Syzygium cumini* L ethanolic extract has bacteriocid effect.

Keywords: Ethanolic extract of Syzygium cumini L, growth, Streptococcus mutans

#### Abstract

Streptococcus mutans mempunyai peranan penting sebagai penyebab karies. Bakteri ini memiliki sifat virulensi yang terlibat dalam pembentukan biofilm kariogenik pada permukaan gigi. Daun Jamblang/Syzygium cumini L mempunyai efek antibakteri sehingga dapat digunakan sebagai bahan untuk mencegah karies. Penelitian ini bertujuan untuk mengetahui efek ekstrak etanolik Syzygium cumini L terhadap pertumbuhan S. mutans. Analisa senyawa aktif dilakukan dengan kromatografi lapis tipis (KLT) dan Liquid Chromatograph-Mass Spectrography (LC-MS). Streptococcus mutans diisolasi dari gigi molar sulung anak yang karies. Uji pertumbuhan dilakukan dengan teknik dilusi. Akuades digunakan sebagai kontrol negatif. Hasil analisis KLT dan LC-MS menunjukkan adanya kandungan flavonoid tannin, dan terpenoid. Uji Kruskall-Wallis menunjukkan adanya perbedaan yang signifikan (p<0,05) antara semua kelompok yang diuji, mengindikasikan ekstrak etanolik Syzygium cumini L menurunkan pertumbuhan S.mutans. Semakin tinggi konsentrasi ekstrak, semakin sedikit jumlah koloni S. mutans yang tumbuh. Pada konsentrasi 22,5% tidak ditemukan lagi pertumbuhan S. mutans. Efek bakteriosid didapatkan pada ekstrak konsentrasi 22,5%.

Kata kunci: Ekstrak etanol Syzygium cumini L, pertumbuhan, Streptococcus mutans

#### INTRODUCTION

Caries is a bacterial infection disease that causes destruction of the hard tissue of teeth. This disease occurs as the result of interactions between teeth, saliva, cariogenic bacteria, food substrate, and time.<sup>1</sup>

Caries is still a major dental and oral problem in children.<sup>2</sup> Children aged 4-6 years (preschool age) are an age group that is in desperate need of attention in dental care. This is because children of that age often consume sweet food, and their ability to maintain oral hygiene is still lacking.<sup>3</sup> One of the pathogenic bacteria that plays an important role in the pathogenesis of caries is Streptococcus mutans (S. Mutans).<sup>1,2</sup> Streptococcus mutans has virulence factors that cause these bacteria to play a role in the occurence of caries. One of the factors is the ability to synthesize and secrete glucosyltransferase (gtf) enzymes.<sup>4,5</sup> Glucosyltransferase consists of gtfB, gtfC, and gtfD. Glucosyltransferase B and gtf C produce glucan bond of  $\alpha$  (1-3) from sucrose, which are very sticky and insoluble in water. Glucans play an important role for attachment and biofilm accumulation.<sup>4,6</sup> Streptococcus mutans has a surface protein peptide antigen (SpaP), namely B antigen (AgB) and I/II antigen (AgI/II) which are the surface proteins of S. mutans. This protein functions to mediate S. Mu*tans* attachment to the tooth surface.<sup>4.7</sup>

Syzygium cumini L is a wild plant that is widely used for health.<sup>8,9</sup> Syzygium cumini L has antimicrobial effects on several bacteria and fungi, including Staphylococcus aureus, Bacillus subtilis, Sa-Imonella typhi, Pseudomonas aeruginosa, Escherichia colli, Candida albicans dan Candida crusei.<sup>9,10</sup> Syzygium cumini L contains a mixture of polyphenols, especially flavonoid glycosides and tannins. In addition, it also contains terpenoid compounds.10,11 Plants that contain flavonoids are known to have antibacterial effects.<sup>12</sup> This study aims to determine the effect of Syzygium cumini L ethanolic extract on the growth of S. mutans that is isolated from dental caries of preschool children (4-6 years), a follow-up study from previous studies using S. mutans OMZ 175.<sup>13</sup> Streptococcus mutans that is used in this study was isolated from children's dental caries because the OMZ 175 Streptococcus mutans that was used in the preliminary study came from humans, but no information was found whether it came from caries or normal lesions, and from children or adults. Therefore, to confirm the test, the S. mutans which was isolated from child caries was used.

## MATERIAL AND METHOD

The research method used is experimental laboratory. The research subjects were kindergarten students in Prujakan, Ngaglik, Sleman. The subjects selected met the criteria of 4-6 years old, suffering from caries that reached dentine in primary molars. Sampling was done by taking soft dentine on carious lesions, using excavators and growing it on *Trypticase-soy yeast extract 20% sucrose with bacitracin* (TYS20B).<sup>14</sup> Research has obtained permission from the Ethics and Advocacy Unit of the Faculty of Dentistry, Gadjah Mada University (FKG UGM).

Syzygium cumini L was taken from Nggembongan village, Sentolo sub-district, Kulon Progo regency, Yogyakarta. Syzygium cumini L which has been cleaned with water, is finely pollinated using a grinder. Syzygium cumini L extract was obtained by maceration using 70% ethanol and diluted with 20% DMSO. The active compound content of Syzygium cumini L extract was analyzed using TLC at UGM Integrated Research and Testing Laboratory (LPPT). The analysis procedure using LC-MS was conducted at the Chemical Research Center Laboratory, Indonesian Institute of Sciences (LIPI), Serpong, West Java.

Streptococcus mutans that was grown on TYS20B media, was put in a wax lid, and incubated for 2x24 hours at 370C.<sup>14</sup> The morphology of S. mutans colonies on TYS20B media gives a round-shape image, 0.5-2µm diameter, and bright white color. In Gram coloring gives Gram positive results with morphology of round cells and cell arrangements forming chains. S. mutans colonies were then cultured on blood agar and in liquid BHI with concentration of 10°CFU/ml. The identification of S. mutans was done by looking at the shape of the colony and the description of hemolysis in blood agar, catalase test, Optochin test, tolerance to 6.5% NaCl, mannitol, sorbitol, inulin and amylum.<sup>15</sup> Test of the effect of Syzygium cumini L ethanolic extract on S. mutans growth was carried out by dilution method. As much as one ml of Syzygium cumini L ethanolic extract was made in three concentrations, i.e 30%, 40%, and 45%. The extract was diluted using 20% DMSO. Then 1 ml of bacterial suspension was added in BHI  $(10^6 \text{ CFU/ml})$ , so the final concentration of the extract was 15%, 20%, 22.5%. Aquades was used as negative controls. All tubes were incubated at  $37^{\circ}C$ for 24 hours. After being diluted in stages using NaCl, then cultured on Muller Hinton Agar (MHA) media. After being incubated for 24 hours at  $37^{\circ}$ C. the number of colonies of bacteria that grow were counted.

#### RESULT

The results of TLC showed the presence of flavornoid active compounds, mainly quercetin (16.6%), tannin (0.19%), and terpenoids. The results of the analysis and identification of compounds in *Syzygium cumini L* ethanolic extract using LC-MS showed the presence of flavonoid compounds, namely *myericetin, 3-0-a-L-rhamnocyl myricetin,* and *taxifolin,* also found terpenoid substance i.e *eugenyl acetate,* and *tricosanoyl lupeol.*  There was a decrease in the number of *S. mutans* colonies after being exposed to *Syzygium cumini L.* etha-nolic extract. The effect of this decrease in-

creased in accordance with the increase in extract concentration. The number of *S. mutans* was found mostly in negative controls (Table 1).

Table 1. Number of S. mutans colonies (CFU/ml) in various concentrations of Syzygium cumini L ethanolic extract

Group	$X \pm SD$
Concentration of extract 22.5%	$0.00 \pm 0.00$
Concentration of extract 20 %	$4866.67 \pm 1.13 \ge 10^3$
Concentration of extract 15%	$5.16 \ge 10^4 \pm 2.44 \ge 10^3$
Negative control	$1.82 \text{ x } 10^{12} \pm 9.99 \text{ x } 10^{10}$

The results of the normality test using *Kosmogorof-Smirnov* showed that the data tested were normally distributed. The homogeneity test shows the data tested is not homogeneous so that the parametric test of variance analysis (ANOVA) cannot be done. The test used was the *Kruskal-Wallis* non-parametric test.

The results of the *Kruskall-Wallis* test showed a significant difference (p < 0.05) in the number of *S*.

*mutans* colonies between groups. These results indicate that exposure to *Syzygium cumini L* ethanolic extract affects the number of *S. mutans* colonies. *Mann-Whitney* test results show a significant difference in the number of *S. mutans* colonies (p <0.05) among all extract concentrations tested (Table 2).

Table 2. Summary of Mann-Whitney test results of *S. mutans* colonies between groups of *Syzygium cumini* L ethanolic extract concentrations

	Between groups		L	Significance
1.	Extract 22.5	Extract 20%	-4.992	0.000*
		Extract 15%	-4.992	0.000*
		Negative control	-4.989	0.000*
2.	20%	Extract 15%	-4.674	0.000*
		Negative control	-4.671	0.000*
3.	15%	Negative control	-4.671	0.000*

Note: \* = significant different (p<0.05)

## DISCUSSION

The results of this study indicate that the ethanolic extract of Syzygium cumini L was able to reduce the number of S. mutans colonies. The higher the extract concentration, the greater the growth barriers that occur. This is due to the components of antibacterial substances in the extract, namely flavonoids, tannins and terpenoids. Increased extract concentration results in a greater decrease in growth. This is related to the increase in the amount of active compounds from *Syzygium cumini L*.<sup>16,17</sup> Flavonoids in *Syzygium* cumini L extract can interfere with the activity of transpeptidase peptidoglycan which disrupts the formation of cell walls, resulting in cell lysis. Flavonoids also form complex compounds with extracellular proteins, thus disrupting S. mutans cell membrane integrity. The disruption of cell membrane integrity, will inhibit protein synthesis or nucleic acid metabolism, inhibit enzyme activity, and inhibit energy metabolism.<sup>18</sup> Inhibition of energy metabolism by flavonoid compounds by inhibiting the accumulation of intracellular polysaccharides (IPS) from S. mutans, so that the function of IPS as a source of endogenous carbohydrates is disrupted. This condition causes inhibition of *S. mutans* growth.<sup>19</sup>

Flavonoids compound of Syzygium cumini L namely mirisetin and quercetin inhibit nucleic acid synthesis. The inhibitory effect of nucleic acid synthesis is related to the presence of ring B from flavornoids. Ring B of flavonoids plays a role in the intercalation or hydrogen bonding with nucleic acids.<sup>12</sup> Syzygium cumini L ethanolic extract in this study contained quercetin. Syzygium cumini L ethanolic extract in this study contained quercetin. Quercetin compounds can inhibit the activity of the enzyme deoxyribonucleic acid girase (DNA girase) and Adenosine triphosphatase (ATPase). The AT Pase enzyme acts to pump protons  $(H^+)$  from the cytoplasm out of the cell, so that the cytoplasmic pH remains in physiological state. If the activity of AT Pase enzyme is inhibited, the cytoplasmic pH becomes acidic, and affects the aciduric nature of S.

*mutans*, as well as *S. mutans* metabolism as a whole. DNA gyrase enzymes function to open the twists of the DNA chain, so as to reduce the temporary tension of double-stranded DNA. This DNA gyrase enzyme is often the target of antibiotics.<sup>4,7</sup> The antibacterial mechanism of tannin compounds is similar to flavonoids, which interfere with membrane lipid and protein interfaces.<sup>20</sup> Tannin compounds can damage bacterial cell membranes, induce the formation of bonding complexes to microbial enzymes or substrates. Tannin compounds can also be toxic to bacteria.<sup>21</sup>

Terpenoids contained in the ethanolic extract of *Syzygium cumini L* can provide antibacterial effects against *S. mutans* through the interaction of lipophilic terpenoid compounds with lipid elements from *S. mutans* cell membrane. This interaction causes disruption of the *S. mutans* structure and makes it more permeable.<sup>22</sup> The terpenoids have membranotropic properties which interfere with the stability of the membrane functional integrity.<sup>23</sup> The terpenoid compounds in 22.5% *Syzygium cumini L* 

#### REFERENCES

4.

- 1. Shivakumar KM, Vidya SK, Chandu GN. Dental caries vaccine. Indian J Dent Res 2009; 20 (1): 99-106.
- 2. Ramos-Gomez F, Crystal YO, Ng MN, Tinanoff, N, John D, Featherstone JD. Caries risk assessment, prevention, and management in pediatric dental care. Gen Dent 2010: 505-17.
- 3. Twetman S, Fontana M. Patient caries risk assessment, dalam detection, assessment diagnosis and monitoring of caries. Monogr Oral Sci 2009; 21: 91-101.
  - uivey RG. Caries, In: Lamont RJ, Burne RA, Lantz MS, Leblanc, eds. Oral microbiology and immunologi., Washington: DJ ASM Press, 2006: 233-52.
- Al-Hebshi NN, Nielsen O, Skaug N. In vitro effects of crude khat extracts on the growth, colonization, and glucosyltransferases of Streptococcus mutans. Acta Odontol Scand 2005; 63: 136-42.
- alaro KP. Foundations in microbiology. Basic principles. 6<sup>th</sup> ed., New York: Mc. Graw Hill Higher Education, 2008: 386.
- Lamont RJ, Jenkinson HS. Oral microbiology at A Glance., Singapore: Willey-Blackwell, 2010: 30-9, 41.
- 8. Kumar A, Ilavarasan R, Jayachandran T, Deecaraman M, Aravindan P, Padmanabhan N, et al. Antidiabetic activity of Syzygium cumini and its isolated

ethanolic extract in this study are likely to be able to lyse membranes or coagulate cytoplasm from S. mutans cells, thus stopping S. mutans growth.<sup>22</sup> This condition is in accordance with the research of Akiyama et al (2001) which proved that, antimicrobial materials that are bacteriostatic can be bacteriocid if used in high dosage.<sup>24</sup> Carvone terpenoid compounds in Syzygium cumini L extract can interfere with the pH gradient and cell membrane potential, thus disrupting the general metabolism of cells. Terpenoid compounds can be bacteriostatic and bacteriocide, depending on their concentration.<sup>19</sup> In this study, terpenoid compounds in Syzygium cumini L extract were bacteriostatic at a concentration of  $\leq 20\%$ , and were bacteriocid at a concentration of 22.5%. The conclusion of this study is that the Syzygium cumini L ethanolic extract has the ability to decrease the growth of S. mutans. Bacteriosid effect is found in the extract concentration of 22.5%. Further study is required to isolate the active substance of Syzygium cumini L so as to understand more in details the active compound that roles in the mechanism of growth inhibition.

compound against Streptozotocin-induced diabetic rats. J Med Plant Res 2008; 2(9):246-9.

9.

e Oliveira GF, Furtado NAJC, Filho AAS, Martins CHG, Bastos JK, Cunha WR, et al. Antibacterial activity of Syzygium cumini (Myartaceae) leaves extract. Braz J Microbiol 2007; 38: 381-4.

- Gowri SS, Vasantha K. Phytochemical screening and antibacterial activity of Syzygium cumini (L.) (Myrtaceae) leaves extracts. Int J Pharm Tech Res 2010; 2(2):1569-73.
- 11. Q Jai n A, Sharma S, Goyal M, Dubey S, Jain S, Sahu J, et al. Anti-inflammatory activity of Syzygium cumini leaves. Int J Phytomed 2010; 2: 124-6.
- Cu shnie TPT, Lamb AJ. Antimicrobial Activity of Flavonoids. Int J Antimicrob Ag 2005; 26: 343-56.
- Sungkar S. Aktivitas Antibakteri Ekstrak Daun Jamblang (Syzigium cumini (L) skeel) terhadap Streptococcus mutans OMZ 175. Th: Indonesian Dental Association, ed. Proceeding Medan International Scientific Dental Meeting. Medan, 2017.
- 14. W a AKL, Seow WK, Walsh LJ, Bird PS. Compa-rison of five selective media for the growth and enumeration of Streptococcus mutans. Aust Dent J 2002; 47(1): 21-6.
- M urray PR, Baron EJ, Jorgensen JH, Pfaller MA, Yolken RH. Manual of Clinical Microbiology, 8th

ed., Washington DC: ASM Press, 2003: 407, 409, 411, 413-5.

- Fe rrazzano GF, Amato I, Ingenito A, Zarrelly A, Pinto G, Pollio A. Plant polyphenols and their anticariogenic properties: A Review. J Mol 2011; 16: 1486-1507.
- Pe lezar MJ, Chan ECS. Dasar-dasar microbiologi. Alih bahasa. Hadioetomo RS. Jakarta: UI press, 2005: 453-6.
- Sil va NCC, Fernandes-Junior A. Biological pro-perties of medicinal plants: A review of their antimicrobial activity. J Venom Anim and Toxins 2010; 16(3): 402-13.
- 19. Je on JG, Rosalen PL, Falsetta ML, Koo H. Natural products in caries research: Current (limited) knowledge, challenges and future perspective. Caries Res 2011; 45: 243–63.
- 20. Gr eenberg M, Dodds M, Tian M. Naturally occur-ring phenolic antibacterial compounds show effecttiveness against oral bacteria by a quantitative struc-

ture-activity relationship study. J Agric Food Chem 2008; 56: 11151-6.

- 21. Sh er A. Antimicroba activity of natural products from medicinal plants. J Gomal Med Sci 2009; 7(1): 72-8.
- Ja wetz E, Menick JL, Adelberg EA. Mikrobiologi kedokteran. ed. 23. Alih bahasa. Eddy Mudihardi. Jakarta: Penerbit Buku Kedokteran EGC, 2008: 23, 225, 229.
- Bu rt S. Essential oil: their antibacterial properties and potential application in food- a review. Int J Food Microbiol 2004; 94(3): 233-53.

24.

A kiyama H, Fujii K, Yamasaki O, Oono T, Iwatsuki T. Antibacterial action of several tannins against Staphylococcusaureus. J Antimicrob Chemother 2001; 48: 487-91.